

Unexpectedly high false-positive rates for *Haemophilus influenzae* using a meningoencephalitis syndromic PCR panel in two tertiary centers

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The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest

Author contribution statement

MCZ, AE, JS contributed to the conception of the work.
MCZ, VH, AC, GR contributed to the data collection.
MCZ, AC, VH, DG, AE, JS contributed to data analysis and interpretation.
MCZ wrote the article.
AG, JS contributed to the critical revision of the article.
MCZ, AC, VH, GR, DG, AE, JS approved the final version to be published.

Keywords

Haemophilus influenzae, False-positive, panel, diagnostics, Meningitis, central nervous system infection and inflammation

Abstract

Word count: 231

False-positive results in the diagnostic of meningitis and encephalitis pose important challenges. This study aimed to determine false-positive rates for Haemophilus influenzae in cerebrospinal fluids evaluated by the BioFire FilmArray® Meningitis/Encephalitis Panel. We conducted a retrospective study of all H. influenzae-positive FilmArray® Meningitis/Encephalitis Panel results from June 2016 to October 2019 in two Swiss university hospitals. Cases were classified as true positive, likely true-positive and likely false-positive results according to cerebrospinal fluid culture, H. influenzae-specific quantitative real-time PCR (qPCR), and Gram staining, as well as culture of other materials. We performed 3,082 panels corresponding to 2,895 patients: 0.6% of samples (18/3,082) were positive for H. influenzae. Culture and H. influenzae-specific qPCR were performed on 17/18 (94.4%) and 3/18 (16.7%) cerebrospinal fluid samples, respectively; qPCR was negative in all cases. Among 17 samples sent for culture, 10 concerned patients not treated with antibiotics prior to lumbar puncture. Only 1/17 revealed growth of H. influenzae and was classified as a true positive. We further classified 3/18 (16.7%) cases with the identification of Gram-negative rods in the cerebrospinal fluid or positive blood cultures for H. influenzae as likely true-positive and 14/18 (77.8%) cases as likely false-positive. Diagnostic results should always be interpreted together with the clinical presentation, cerebrospinal fluid analysis and other available microbiological results. All H. influenzae-positive results should be viewed with special caution and a H. influenzae-specific qPCR should be systematically considered.

Contribution to the field

False-positive results in the diagnosis of meningitis and encephalitis cause important challenges. Several cases of false-positive results for some targeted pathogens, and more specifically for Haemophilus influenzae, have been reported with the BioFire FilmArray® ME Panel. We conducted a retrospective study of all H. influenzae-positive FilmArray® ME Panel results on cerebrospinal fluid (CSF) samples, over a period of 3 years at Geneva and Basel university hospitals. Cases were classified as true positive, likely true-positive and likely false-positive results according to microbiological results, namely CSF culture, H. influenzae-specific qPCR, Gram staining, and culture of other specimens. Our study revealed H. influenzae BioFire FilmArray® ME Panel positive results in 0.6% (95% CI 0.4-0.9%) of CSF samples. The review of the 18 H. influenzae-positive results revealed that only 1 (5.5%) case could be considered as a true positive and 3 (16.7%) cases as likely true-positive results. A high proportion of cases (77.8%) were therefore classified as likely false-positive. In addition among these cases, a review of medical charts suggested that H. influenzae-associated meningitis was finally retained in a few patients, with an alternative diagnosis made in 92.9% of patients. In the context of a lack of extensive, well-designed and high-quality analytical and clinical validation studies for syndromic panels, this study may contribute to the surveillance of the above-mentioned assay's performance specifically regarding H. influenzae. Our study highlights the need for caution when reporting H. influenzae-positive results with the BioFire FilmArray® ME Panel. Importantly, such results should always be interpreted together with clinical manifestations, CSF analysis and other microbiological results.

Ethics statements

Studies involving animal subjects

Generated Statement: No animal studies are presented in this manuscript.

Studies involving human subjects

Generated Statement: The studies involving human participants were reviewed and approved by Geneva and Basel cantonal ethics commissions . Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

Inclusion of identifiable human data

Generated Statement: No potentially identifiable human images or data is presented in this study.

In review

Data availability statement

Generated Statement: The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

In review

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22 **Keywords: *Haemophilus influenzae*, false-positive, panel, diagnostics, meningitis, central**
23 **nervous system infection**

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26

27 **Abstract**

28 False-positive results in the diagnostic of meningitis and encephalitis pose important challenges. This
29 study aimed to determine false-positive rates for *Haemophilus influenzae* in cerebrospinal fluids
30 evaluated by the BioFire FilmArray® Meningitis/Encephalitis Panel. We conducted a retrospective
31 study of all *H. influenzae*-positive FilmArray®. Meningitis/Encephalitis Panel results from June 2016
32 to October 2019 in two Swiss university hospitals. Cases were classified as true positive, likely true-
33 positive and likely false-positive results according to cerebrospinal fluid culture, *H. influenzae*-
34 specific quantitative real-time PCR (qPCR), and Gram staining, as well as culture of other materials.
35 We performed 3,082 panels corresponding to 2,895 patients: 0.6% of samples (18/3,082) were
36 positive for *H. influenzae*. Culture and *H. influenzae*-specific qPCR were performed on 17/18
37 (94.4%) and 3/18 (16.7%) cerebrospinal fluid samples, respectively; qPCR was negative in all cases.
38 Among 17 samples sent for culture, 10 concerned patients not treated with antibiotics prior to lumbar
39 puncture. Only 1/17 revealed growth of *H. influenzae* and was classified as a true positive. We
40 further classified 3/18 (16.7%) cases with the identification of Gram-negative rods in the
41 cerebrospinal fluid or positive blood cultures for *H. influenzae* as likely true-positive and 14/18
42 (77.8%) cases as likely false-positive. Diagnostic results should always be interpreted together with
43 the clinical presentation, cerebrospinal fluid analysis and other available microbiological results. All
44 *H. influenzae*-positive results should be viewed with special caution and a *H. influenzae*-specific
45 qPCR should be systematically considered.

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52 **1 Introduction**

53 Central nervous system (CNS) infections are commonly caused by diverse pathogens, mostly viruses
54 and bacteria and molecular multiplex panels have been developed to simultaneously diagnose
55 multiple pathogens in cerebrospinal fluid (CSF) samples. These molecular rapid diagnostic tests are
56 increasingly used in routine diagnostic laboratories for the diagnosis of CNS infections, including
57 meningitis and meningoencephalitis, with the advantages of being easy to use and having short turn-
58 around times. Nevertheless, these syndromic panels lack extensive analytical and clinical validation
59 and concerns have been raised regarding their suboptimal performances (1). Several cases of false-
60 positive results for some targeted pathogens, more specifically for *Haemophilus influenzae*, have
61 been reported with the BioFire FilmArray® Meningitis/Encephalitis (ME) Panel (bioMérieux, Lyon,
62 France) (2, 3). False-positive results could lead to additional investigations and inappropriate
63 antibiotic prescriptions.

64 *H. influenzae* is associated with upper and lower respiratory tract infections, but also with invasive
65 infections including meningitis (4). The broad use of the *Haemophilus influenzae* type b (Hib)
66 conjugate vaccine led to a dramatic reduction of Hib infections in various countries including
67 Switzerland, where the incidence in 2019 was of 1.43 confirmed case per 100'000 population (4, 5).
68 In Switzerland, the reported vaccination coverage for children is 89% for the 2014-2016 period (6).
69 After implementation of the FilmArray® ME Panel, the bacteriology laboratories of Geneva and
70 Basel university hospitals witnessed an unexpectedly high number of positive cases of *H. influenzae*,
71 raising suspicion of false-positive cases. In this context, the aim of this retrospective study was to
72 analyze all reports of *H. influenzae* and to determine the proportion of false-positive cases on all CSF
73 samples analyzed in these two laboratories starting from the introduction of the FilmArray® ME
74 Panel.

75

76 **2 Materials and methods**

77 **2.1 Ethics statement**

78 The study was approved by the Geneva and Basel cantonal ethics commissions (Geneva #2020-
79 00215; Basel #2020-01724).

80 **2.2 Clinical specimens**

81 We included all samples from pediatric and adult patients for whom a FilmArray® ME Panel PCR
82 (bioMérieux, Lyon, France) was ordered for CSF microbiology testing at the bacteriology
83 laboratories of Geneva and Basel university hospitals from 1 June 2016 (Geneva) and 17 May 2016
84 (Basel) to 31 December 2019 and reported to be positive for *H. influenzae*. Multiple specimens from
85 single patients were not excluded.

87 **2.3 Routine, FilmArray® ME Panel, and confirmatory testing**

88 Both laboratories are accredited (ISO/IEC17025) and regularly participate in external quality control
89 surveys. In both hospitals, all patients with suspected CNS infection benefit from clinical evaluation,
90 prompt administration of antimicrobials, cerebral radiological exams when needed, and lumbar
91 puncture for CSF analysis, according to published guidelines (7). When sufficient CSF sample
92 volume is available, the following analyses are performed: CSF cellular count and chemistry, and
93 direct examination with Gram stain, culture, and the FilmArray® ME Panel. Further microbiological
94 investigations can be performed according to the clinical presentation and suspected pathogen, local
95 epidemiology, in accordance with published guidelines (7). *H. influenzae* positive results with the
96 FilmArray® ME panel are confirmed with a specific quantitative real-time PCR (qPCR) as previously
97 reported (8) when enough CSF sample is available.

98 The FilmArray® ME Panel was performed on CSF samples in accordance with the manufacturer's
99 instructions, including laboratory precautions and good microbiological practice to prevent
100 contamination. Briefly, 200 µl of CSF specimen are subjected to panel testing. The FilmArray® ME
101 Panel test consists of automated nucleic acid extraction, reverse transcription, two multiplex nucleic
102 acid PCR amplifications, and detection using DNA melting temperature analysis of 14 of the most
103 common bacterial, viral and fungal pathogens associated with CNS infections (7 viruses, 6 bacteria
104 and *Cryptococcus neoformans/gattii*) (9); results are available within approximately 1 hour.

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106 **2.4 Data collection**

107 Chart review was conducted to collect the characteristics of the included patients and data regarding
108 their clinical management. During clinical management, the diagnosis of CNS infection was based on
109 the integration of the pre-test probability, clinical presentation, radiological examinations and CSF
110 analysis results. Patients received antimicrobial treatment according to published guidelines (7). A
111 review of laboratory data was conducted to collect CSF cell counts, as well as protein and chemistry
112 values of the included CSF samples. Results of culture and other molecular assays were obtained for
113 the included CSF samples and additional clinical samples collected within 48 hours around the time
114 of CSF sampling. Medical charts were also reviewed for adverse events attributed to antibiotic
115 treatment and *Clostridioides difficile* infection during a maximum of 2 months in-hospital follow-up.

116

117 **2.5 Data analysis**

118 Positive results for *H. influenzae* with the FilmArray® ME Panel and further laboratory tests were
119 classified as follows:

120 - *True-positive result*: confirmed by CSF culture and/or positive *H. influenzae*-specific qPCR on the
121 same CSF sample.

122 - *Likely true-positive result*: not confirmed by CSF culture or *H. influenzae*- specific qPCR when
123 performed. CSF cellularity, chemistry analysis, CSF Gram stain results and the culture results of
124 other clinical samples (e.g. blood cultures) suggest that the result was likely to be a true positive.

125 - *Likely false-positive result*: not confirmed by CSF culture or *H. influenzae*-specific qPCR when
126 performed. CSF cellularity, chemistry analysis, CSF Gram stain results and the culture results of
127 other clinical samples (e.g. blood cultures) suggest that the result was likely to be a false-positive.

128 **3 Results**

129 We performed a total of 3'082 FilmArray® ME Panels corresponding to 2'895 patients (2'252 adult;
130 643 pediatric patients). Results for *H. influenzae* were negative in 99.4% (3'064/3'082; 95% CI 99.1–
131 99.6%) cases and positive in 0.6% (18/3082; 95% CI 0.4–0.9%) CSF samples, corresponding to 12
132 adult and 6 pediatric patients. Clinical characteristics and laboratory data of the 18 cases with
133 positive *H. influenzae* FilmArray® ME Panel results are detailed in Table 1. Among the 18 cases, 5
134 cases (patients #1, #2, #3, #5, and #6, Table 1) and 13 cases had CSF specimens collected and
135 analyzed in Basel and Geneva University Hospital Laboratory, respectively.

136

137 Culture and *H. influenzae*-specific qPCR were performed on 17/18 (94.4%) and 3/18 (16.7%) CSF
138 samples, respectively. *H. influenzae*-specific qPCR assay was performed in 3 samples only due to the
139 limited CSF sample volume available for further testing; qPCR was negative in 3 cases. Among 17
140 samples sent for culture, 10 concerned patients were not treated with antibiotics prior to lumbar
141 puncture and only 1 revealed the growth of *H. influenzae*. This case (patient #1, Table 1) was
142 classified as a true positive. We classified 3 (16.7%) cases as likely true-positive, with the

143 identification of Gram negative rods on CSF gram stain in 2 cases (patients #2 and #3, Table 1) and
144 positive blood cultures for *H. influenzae* in 2 cases (patients #2 and #4, Table 1). Of note, patients #2
145 and #4 were diagnosed with *H. influenzae* otitis media. The remaining 14 (77.8%) cases were
146 classified as likely false-positive cases (Table 1).

147 CSF white blood cell count was determined in 13 samples (median 7 M/L [range 1-4'031M/L]).
148 Among patients with an elevated CSF white blood cell count, 2 cases were classified as likely true-
149 positive *H. influenzae* results (patients #3 and #4, Table 1), and 2 as likely false-positive with another
150 retained cause of CNS infection (patient #10 with enterovirus-associated meningitis and patient #14
151 with *Escherichia coli*-associated meningitis, Tables 1 and 2).

152
153 Regarding the clinical management, the review of medical charts identified that an alternative
154 diagnosis was made for 13/18 (72.2%) patients (Table 2). All cases classified as likely true- positive
155 were treated for *H. influenzae* meningitis (Table 2). Among cases classified as likely false-positive,
156 only 4 were treated with antibiotics for bacterial meningitis (Table 2). No adverse event associated
157 with antibiotic treatment or *Clostridioides difficile* infection were reported (Table 2).

158
159 Details of the amplification curve results of the initial and repeated FilmArray® ME Panels when
160 performed are provided in Table S1. Only 1/18 (5.5%) samples revealed positive amplification
161 curves on the two targets designed to detect *H. influenzae* (patient #4, Table S1). Among 3 patients
162 for whom the FilmArray® ME Panel was repeated on the same CSF sample, results were positive in 2
163 patients and classified as likely true-positive cases (patients #2 and #4, Table S1).

164

165 4 Discussion

166 In this two-center retrospective study conducted over a period of 3 years, the proportion of CSF
167 samples with *H. influenzae* FilmArray® ME Panel positive results was 0.6% (95% CI 0.4–0.9%). A
168 review of the 18 *H. influenzae* positive results revealed that only 1 case (5.5%) could be considered
169 as a true positive. Regarding *H. influenzae* culture-positive results from other clinical samples, Gram
170 stain and CSF analysis, 16.7% of cases were classified as likely true-positive cases. At the time of
171 clinical management, these patients have been treated accordingly. For a high proportion of cases
172 (77.8%), CSF analysis and microbiological results suggested that *H. influenzae* FilmArray® ME
173 positive results were likely false-positive. Among these, the review of medical charts suggested that
174 *H. influenzae*-associated meningitis was finally retained in a few patients : an alternative diagnosis
175 was made in 92.9% of patients and only 28.6% received a full course of antibiotics for bacterial
176 meningitis. The fact that only a minority received antibiotics highlights that the result was not in
177 concordance with the clinical picture and course of the patient. The impact of the false-positive
178 results was therefore limited since panel results were interpreted together with clinical
179 manifestations, CSF analysis and other microbiological results. Importantly however, neither the
180 validation studies performed before panel implementation nor the regular EQC are designed to detect
181 that rate of false-positive results, thus prompting such reports by consortia of panel users.

182 In the context of a lack of extensive analytical and clinical validation studies, this study may
183 contribute to the surveillance of the assay performance specifically regarding *H. influenzae*. Among
184 the *H. influenzae* false-positive results reported in the literature (2, 3), it remains unclear whether the
185 false-positive results are due to pre-analytic issues, sample contamination or to an analytical artifact.
186 However, in our study, sample contamination seems unlikely given the rigorous handling of the
187 samples, the analyses performed in two different laboratories, and the absence of frequent false-
188 positive results for *Streptococcus pneumoniae* for instance, which could be at least as frequent as an
189 external contaminant; qPCR results revealed to be all negative, thus suggesting, at least in the tested

190 samples, that the positive results of the *H. influenzae* FilmArray® ME Panel were likely false-
191 positive. False-positive results highlight the precautions needed to perform rapid diagnostic tests to
192 avoid cross-contamination from the environment and especially from the personnel in contact with
193 the sample during the pre-analytical and analytical phases of testing.

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195 Of note, there is also a lack of validation studies assessing the performance of the assay for each
196 pathogen individually. Specifically regarding *H. influenzae*, the sensitivity of the FilmArray® ME
197 Panel compared to culture may in fact confer an advantage for CNS infection diagnosis, as reported
198 in a recent study (10). Nevertheless, our study raises the issue of its potential lack of specificity for
199 *H. influenzae*, as reported by others (2, 3). Globally, the conclusions drawn on the basis of the
200 performances of the assay based on the few validation studies available are notably limited by the
201 low number of samples tested for each pathogen.

202

203 Our study has some limitations. Specific qPCR was not systematically performed in the context of an
204 insufficient available CSF sample. Nevertheless, qPCR results revealed to be all negative, thus
205 suggesting, at least in the samples tested, that the positive results of the *H. influenzae* FilmArray®
206 ME Panel were likely false-positive. Some laboratory analyses were missing, precluding a rigorous
207 classification of some cases. The retrospective review of clinical charts did not identify any antibiotic
208 adverse events or *C. difficile* infections, although we cannot exclude information bias due to the lack
209 of reporting or the occurrence of such events after hospitalization.

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211 In conclusion, this study highlights the need for caution for *H. influenzae*-positive results with the
212 FilmArray® ME Panel. Meningoencephalitis syndromic panel results should always be interpreted

213 together with clinical manifestations, CSF analysis and other microbiological results. The use of a
214 specific *H. influenzae* qPCR on CSF samples should be systematically considered as a confirmatory
215 assay.

216

217 **5 Conflict of Interest**

218 *The authors declare that the research was conducted in the absence of any commercial or financial*
219 *relationships that could be construed as a potential conflict of interest.*

220

221 **6 Author Contributions**

222 MCZ, AE, JS contributed to the conception of the work

223 MCZ, VH, AC, GR contributed to the data collection

224 MCZ, AC, VH, DG, AE, JS contributed to data analysis and interpretation

225 MCZ wrote the article

226 AG, JS contributed to the critical revision of the article

227 MCZ, AC, VH, GR, DG, AE, JS approved the final version to be published

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231

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281 **9 Tables**

282 **Table 1:** Case classification, patient characteristics and laboratory data of patients with positive *H.*
283 *influenzae* BioFire FilmArray® ME Panel results (n=18).

284 **Table 2:** Alternative diagnosis, clinical management and reported adverse effects of antibiotic
285 treatment based on the retrospective review of medical charts of patients with positive *H. influenzae*
286 BioFire FilmArray® ME Panel results (n=18).

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288 **10 Supplementary Material**

289 **Table S1:** Detailed results of the BioFire FilmArray® ME Panel for positive *H. influenzae*
290 cerebrospinal fluid samples (n=18).

291

292 **Data Availability Statement**

293 The original contributions presented in the study are included in the article/supplementary materials,
294 further inquiries can be directed to the corresponding author/s.

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304 **Table 1:** Case classification, patient characteristics and laboratory data of patients with positive *H. influenzae* BioFire FilmArray® ME Panel
 305 results (n=18).

Patient number	Case classification*	Gender	Age (years)	Antibiotics <48 h before LP	CSF and blood chemistry					CSF microbiological testing			Other suspected infection sites	Blood culture results**	Other clinical sample culture results	
					CSF WBC (M/L)	CSF PMN (%)	CSF total proteins (g/L)	CSF glucose (mmol/L)	Blood glucose (mmol/l)	CSF Gram stain	CSF culture	Biofire FilmArray® ME Panel result				Specific <i>H. influenzae</i> qPCR
1	TP	M	5	NA	NA	NA	NA	NA	NA	Gram neg rods	<i>H. influenzae</i>	<i>H. influenzae</i>	ND	no	neg (1/1)	
2	Likely TP	M	1	no	NA	NA	NA	NA	NA	Gram neg rods	neg	<i>H. influenzae</i>	ND	Otitis media	<i>H. influenzae</i> (1/1)	Ear-swab, <i>H. influenzae</i>
3	Likely TP	M	21	yes	90	65	0.81	3.5	ND	Gram neg rods	neg	<i>H. influenzae</i>	ND	no	neg (4/4)	
4	Likely TP	F	2	no	4031	78	0.77	3.9	7.6	absence	neg	<i>H. influenzae</i>	ND	Otitis media	<i>H. influenzae</i> (1/1)	
5	Likely FP	M	79	no	5	0	1.22	9.1	ND	absence	neg	<i>H. influenzae</i>	ND	no	neg (4/4)	
6	Likely FP	M	11	no	NA	NA	0.99	2	ND	Gram pos cocci	<i>S.aureus</i>	<i>H. influenzae</i>	ND	Ventricular shunt infection	neg (1/1)	
7	Likely FP	M	12	yes	3	58	0.85	2.8	3.7	absence	neg	<i>H. influenzae</i>	neg	no	ND	
8	Likely FP	F	34	yes	1	54	0.45	3.9	6.3	absence	neg	<i>H. influenzae</i>	ND	no	neg (4/4)	
9	Likely FP	F	1 month	no	7	20	0.65	2.5	ND	absence	neg	<i>H. influenzae</i>	neg	no	<i>Actinomyces</i> sp.† (1/1)	
10	Likely FP	F	26	yes	230	10	0.71	3.4	ND	absence	neg	<i>H. influenzae</i> , enterovirus	neg	no	neg (4/4)	
11	Likely FP	M	49	yes	2	1	1.03	4.3	5.9	ND	ND	<i>H. influenzae</i>	ND	no	neg (4/4)	
12	Likely FP	F	22	yes	<1	ND	0.28	3.8	5.7	absence	neg	<i>H. influenzae</i>	ND	no	ND	
13	Likely FP	M	82	no	4	18	0.86	3.7	5.4	absence	neg	<i>H. influenzae</i>	ND	no	neg (2/2)	
14	Likely FP	M	77	no	3623	98	3.89	7.8	13.2	Gram neg rods	<i>E. coli</i> K1	<i>H. influenzae</i> , <i>E. coli</i> K1	ND	no	neg (6/6)	
15	Likely FP	F	46	no	<3	0	0.29	ND	ND	absence	neg	<i>H. influenzae</i>	ND	no	ND	
16	Likely FP	F	73	yes	2	9	0.31	5.4	5.1	absence	neg	<i>H. influenzae</i>	ND	no	ND	
17	Likely FP	M	38	no	14	12	0.66	3.2	ND	absence	neg	<i>H. influenzae</i>	ND	no	ND	
18	Likely FP	F	43	no	9	5	0.39	3.4	5.1	absence	neg	<i>H. influenzae</i>	ND	no	ND	

306 *See Materials and Methods section for case classification definitions.

307 ** Number of positive or negative blood cultures bottles among total blood cultures sampled within 48 h around the time of the lumbar puncture.

308 † Considered as a contaminant.

***H. influenzae* false-positive rates using a syndromic PCR panel**

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Abbreviations: FP: false-positive; TP: true-positive; M: male; F: female; LP: lumbar puncture; CSF: cerebrospinal fluid; qPCR: quantitative real-time PCR; WBC: white blood cell count; Lympho: lymphocytes; PMN: polymorphonuclear; *H. infl.*: *H. influenzae*; *S. aureus*: *Staphylococcus aureus*; *E. coli*: *Escherichia coli*; neg: negative; pos: positive; ND: not done; NA: not available.

In review

H. influenzae false-positive rates using a syndromic PCR panel

329 **Table 2:** Alternative diagnosis, clinical management and reported adverse effects of antibiotic treatment based on the retrospective review of
330 medical charts of patients with positive *H. influenzae* BioFire FilmArray® ME Panel results (n=18).

Patient number	Case classification*	Alternative diagnosis	Final retained alternative diagnosis	Treated as bacterial meningitis with full course of antibiotics	Reported CDI or adverse event associated with antibiotics
1	TP	no	none	yes	NA
2	Likely TP	no	none	yes	NA
3	Likely TP	no	none	yes	no
4	Likely TP	no	none	yes	no
5	Likely FP	yes	Encephalopathy/encephalitis of unknown etiology	yes	no
6	Likely FP	yes	<i>S. aureus</i> -associated ventricular shunt infection	yes	NA
7	Likely FP	yes	Influenza A-associated meningo-encephalitis	yes	no
8	Likely FP	yes	Epilepsy crisis associated with benzodiazepine withdrawal	no	no
9	Likely FP	yes	Fever of unknown origin	no	no
10	Likely FP	yes	Enterovirus-associated meningitis	no	no
11	Likely FP	yes	Epilepsy crisis associated with cerebral toxoplasmosis and pachymeningitis of unknown origin	no	no
12	Likely FP	yes	Fever and headache (suspicion of upper respiratory tract infection)	no	no
13	Likely FP	yes	Epilepsy and delirium after cranial trauma	no	no
14	Likely FP	yes	<i>E. coli</i> K1-associated meningitis	yes	no
15	Likely FP	yes	Delirium of unknown origin in a patient with HIV infection	no	no
16	Likely FP	no	none	yes	no
17	Likely FP	yes	Chronic pachymeningitis of unknown origin	yes	no
18	Likely FP	yes	Multiple sclerosis	no	no

331 *See Material and Methods section for case classification definitions

332 *Abbreviations:* FP: false positive; TP: true positive; NA: not available. CDI : *Clostridioides difficile* infection. *S. aureus*: *Staphylococcus aureus*. *E. coli* : *Escherichia coli*.

In review

In review